

found in great numbers in Roman deposits at several sites in the city, yet are absent from 8th to 11th century deposits, and reappear in the fossil record in the 12th to 13th century and later levels.

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REFERENCES

- Armitage, P., West, B. & Steedman, K. (1984). New evidence of black rat in Roman London. *London Archaeol.* **4**(14): 375–383.
- Corbet, G. B. & Southern, H. N. (Eds) (1977). *The handbook of British mammals*. 2nd edn. Oxford: Blackwell.
- O'Connor, T. P. (1988). Bones from the General Accident site, Tanner Row. *Archaeology of York* **15**(2): 61–136.
- O'Connor, T. P. (1989). Bones from Anglo-Scandinavian levels at 16–22 Coppergate. *Archaeology of York* **15**(3): 137–208.
- Rackham, D. J. (1979). *Rattus rattus*. The introduction of the black rat into Britain. *Antiquity* **53**: 112–120.

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Identification of fish otoliths and bones in faeces and digestive tracts of seals

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Introduction

Diet analysis for seals is normally based on visual recognition of prey remains in digestive tracts or faeces. Thus, studies by Rae (1960, 1968, 1973) were based on identification of hard remains of fish and invertebrates in stomach contents. More recent work has utilized identification and measurement of otoliths in faeces (e.g. McConnell *et al.*, 1984; Prime & Hammond, 1985; Härkönen, 1987, 1988; S.M.R.U., 1988).

Faecal and digestive tract samples present different kinds of problems in relation to the logistics of sample collection, the identification of prey remains, and the quantification of diet composition.

Faeces can only be collected on land, e.g. at breeding and haul-out sites, and sites may not be used by seals all year round (e.g. Thompson, 1989). Intertidal haul-out sites can be sampled only at low tide. Digestive tract samples can be collected at sea, but carcasses of pinnipeds often sink; e.g. two-thirds of sea-lions killed at sea by Fiscus & Baines (1966) sank and could not be recovered. Another widely reported problem is that stomachs of seals killed on land are often empty (e.g. Pikharev, 1946; Kenyon, 1956), although the extent of this problem will vary seasonally, e.g. because some species fast during the breeding season. There are obviously also aesthetic, legal and moral issues associated with killing seals. Lavaging of stomach contents (e.g. Antonelis *et al.*, 1987) potentially offers a non-destructive alternative.

The ease with which prey remains in digestive tract and faecal samples can be identified depends on the extent to which they have been degraded during ingestion and digestion. Only hard remains (e.g. fish otoliths and bones, cephalopod beaks) are likely to be identifiable in faeces; however,

some hard parts are entirely digested and many are reduced in size during passage through the digestive tract (e.g. da Silva & Neilson, 1985; Murie & Lavigne, 1986; Jobling & Breiby, 1986; Prime & Hammond, 1987; Harvey, 1989). Differential passage rates are a potential source of bias when diet is assessed from stomach contents (Bigg & Fawcett, 1985; Prime & Hammond, 1987). Although many studies of diet have involved identification of fish bones (e.g. Hansel *et al.*, 1988), the extent to which this increases the amount of information obtained from samples has not been evaluated.

There are several methods of quantifying diet composition, e.g. frequency of occurrence, numbers of prey, prey biomass: see Bigg & Perez (1985) for a discussion of the biases associated with each. Estimation of prey biomass is normally based on measurements of fish otoliths and cephalopod beaks, corrected for reduction in size during passage through the digestive tract (e.g. Prime & Hammond, 1987). However, some authors consider that the errors involved in this approach are so great that otoliths in faeces should not be used as the basis for quantification of diet (Jobling & Breiby, 1986; Jobling, 1987).

In the present paper, digestive tract and faecal samples, from common seals (*Phoca vitulina*) and grey seals (*Halichoerus grypus*), collected in the course of a three-year study of the diets of seals in Scottish waters, are used to evaluate aspects of methodology. Specifically, we consider the logistics of collecting faecal samples, the proportion of digestive tract and faecal samples which contained identifiable prey remains, and the extent to which use of skeletal elements other than otoliths enhanced the information obtained from samples.

Methods

Digestive tract samples were supplied by the Department of Agriculture and Fisheries for Scotland (see Table I for details). In all cases, species was determined by examining the head or lower jaw (Corbet & Southern, 1977). The digestive tracts were stored at -20°C prior to examination. Anecdotal information was obtained from fishermen on the success of attempts at recovering seals which had been shot.

TABLE I
Origin of digestive tract samples. All seals were taken from the east coast of Scotland between Helmsdale and Arbroath

Source	Number of samples	
	Common seals	Grey seals
Fishermen (in or near salmon nets)	6	23 ¹
Fishermen (away from nets)	4	2
DAFS* (taken for tissue samples)	0	15
Found on beaches	0	3 ²
Total numbers of samples	10	43

* Department of Agriculture & Fisheries for Scotland

¹ Includes two samples of stomachs only

² Two of these animals were found injured and put down by vets

TABLE II

Origins of faecal samples. Numbers of common seal scats (C), grey seal scats (G) and numbers of scats of unknown origin (U) collected during each quarter of the year in each area, over the period June 1986 to February 1989. ('—' indicates that the site was not visited.)

Season	Jan.-Mar.			Apr.-Jun.			Jul.-Sep.			Oct.-Dec.			Totals		
Location	C	G	U	C	G	U	C	G	U	C	G	U	C	G	U
Helmsdale	—	—	—	0	21	0	—	—	—	—	—	—	0	21	0
Isle of May	0	17	0	—	—	—	—	—	—	0	75	0	0	92	0
Moray Firth	83	0	1	54	11	36	129	1	31	41	0	0	307	12	68
Orkney	0	21	9	93	0	0	103	0	0	0	35	0	196	56	9
Summer Isles	0	62	0	0	0	0	0	0	0	0	0 ¹	0	0	62	0
All areas	83	100	10	147	32	36	232	1	31	41	110	0	503	243	77

¹ Grey seal faeces were collected by DAFS at the Summer Isles in December, but were lost in transit

Faecal samples were collected at haul-out sites around Scotland (see Table II) during the period June 1986 to February 1989. Sites were visited at or around low tide and all faeces found were collected in polythene bags. When possible, seals using the haul-out were counted and identified to species before a collection was made. Samples were stored at -20°C prior to examination.

In the Moray Firth area faeces (primarily from common seals) were sampled during every month of 1988. Haul-out sites in the Beaulay, Cromarty, Inverness and Dornoch Firths were visited by boat or on foot. Sampling by boat was attempted only in good weather conditions, to visit sites known to be used by common seals (Paul Thompson, pers. comm.). On each occasion, it was normally possible to visit sites in one Firth only.

Digestive tract samples were divided by ligatures into stomach, intestine and colon. The contents of each section were squeezed out and washed through a 0.355 mm sieve, to give separate samples for each section. All sections were then washed out into the sieve to recover any material adhering to the mucosa (residual washings). Faecal samples were also washed through a 0.355 mm sieve. All material remaining in the sieve was retained. All otoliths and a sample of bony remains were stored dry in glass vials. Cephalopod beaks and other invertebrate remains were stored in 95% ethanol.

A reference collection of otoliths and skeletons of North Sea fish was established. Between one and six specimens of 67 species were prepared. Flesh was softened by immersion in boiling water or cooking in a microwave oven. The bulk of flesh was removed and bones cleaned by soaking in a saturated solution of Bio-tex (Blumoller Ltd., Denmark). If necessary, fat was removed by soaking in a 50:50 mixture of acetone and chloroform. Skeletal elements were identified and stored dry in individually labelled containers.

Otoliths were identified using the reference collection and published otolith guides (Brodeur, 1979; Breiby, 1985; Härkönen, 1986). Other skeletal elements were identified using the reference collection. Bones which were recognizable to species and occurred in samples included the premaxilla, maxilla, dentary, articular, vomer, operculum, preoperculum, post-temporal, supra-cleithrum, cleithrum, hyal, urohyal, hyomandibular, palatine, otic capsules, teeth and denticles; also the atlas, ultimate and caudal vertebrae (see Ford, 1937; Norden, 1961 for terminology; Mujib, 1967).

TABLE III

Success of faecal sampling in the Moray Firth area: Numbers of successful (i.e. >0 samples) and unsuccessful sampling trips, and average number of samples obtained in each quarter with standard deviation in parentheses

Quarter	Number of visits		Mean number of samples	
	Unsuccessful	Successful	Per visit	Per successful visit
Jan.-Mar.	12	13	3.5 (6.5)	6.8 (7.6)
Apr.-Jun.	6	17	4.5 (4.8)	6.1 (4.7)
Jul.-Sep.	2	19	7.7 (8.0)	8.5 (8.0)
Oct.-Dec.	3	10	3.2 (4.1)	4.1 (4.2)
Totals	23	59	4.8 (6.4)	6.7 (6.7)

Results

Sample collection

Faeces were generally found as discrete units of solid or semi-solid material. In some cases several pieces of material were found along the track left by a seal moving towards the water, and were considered to be part of the same sample. Faecal material at the water's edge tended to have been broken up by wave action and was not included in the analyses.

Of 82 sampling trips in the Moray Firth area, faeces were found on 59 occasions (72%), with an average of 6.7 samples per 'successful' trip. Sampling trips were generally more successful in the summer than in the winter (see Table III). There was a significant positive relationship between the total number of seals present on haul-outs and the number of scats found (Spearman's rank correlation, $r=0.373$, $N=76$ trips, $P<0.01$).

Fishermen who supplied seal carcasses to DAFS reported, on several occasions, that seals shot near salmon nets or haul-out sites could not be recovered but were later found washed ashore. In one instance, of eight seals shot and killed in the water, only one could be recovered before it sank.

Identification of the species of seal which produced a faecal sample depends on information on the animals using a haul-out. Only grey seals were present at haul-outs on the Summer Isles and Isle of May. On Eynhallow in Orkney, both species were present but with different seasonal distributions. Both species are present in the Moray Firth, although grey seals occurred largely in the outer Dornoch Firth. At mixed haul-outs, identification of samples to species was necessarily tentative. Samples were recorded as of unknown origin if no seals were seen or if neither species comprised more than 90% of seals present.

Presence of prey remains in samples

Table IV summarizes information on the number of tracts containing recognizable food remains and the location of food remains in the digestive tracts. Most samples contained fish remains identifiable at least to Order (Table V). In many cases identification to species was possible.

Most faecal samples contained prey remains (Table V). However, up to 20% of grey seal faeces

TABLE IV

Proportion of digestive tracts containing recognizable food remains. Negative results (i.e. the absence of prey remains from a given section) were included only if more than 90% of the otoliths recovered from the whole tract were recovered in separate sectional samples (i.e. < 10% otoliths in residual washings)

Section	Sample size	Containing prey remains
Stomach	44	27 (61.4%)
Intestine	42	29 (69.0%)
Colon	48	43 (89.6%)
Entire tract	51	48 (94.1%)

contained only fragments of fish bones which could not be identified further. Almost 95% of common seal samples contained fish remains identifiable at least to Order (and often to species).

There are two potential sources of error in otolith identification: loss of information due to failure to identify otoliths, and erroneous identification. The maximum loss of information due to failure to identify otoliths is given by the proportion of unidentified otoliths in the samples. In digestive tract samples, 99.7% of otoliths (N = 6587) were identifiable at least to Order. For faecal samples the figures were: 99.9% (N = 50792) for common seal samples, and 98.9% (N = 6753) for grey seal samples. Identification to broad taxonomic groups is considered to be relatively

TABLE V

Categories of prey identification in digestive tract and faecal samples from common and grey seals. Note: fragments of mollusc shell were not counted as prey remains although frequently present, since they may originate from the substratum

Identification category	Digestive tract samples	
	Common seals (N = 10) (%)	Grey seals (N = 41) (%)
Prey present	70.0	100.0
Fish prey present	70.0	97.6
Fish identifiable at least to Order	70.0	90.2
Identification category	Faecal samples	
	Common seals (N = 503) (%)	Grey seals (N = 243) (%)
Prey present	98.2	98.4
Fish prey present	98.2	98.4
Fish identifiable at least to Order	94.6	79.8

TABLE VI

Detection of otoliths and bones in seal digestive tract and faecal samples. Numbers of samples in which (a) otoliths, (b) bones, and (c) otoliths and/or bones of major prey groups were found, and the percentage of cases in which the prey would not have been detected using otoliths alone. Prey types occurring in fewer than five samples are not included in the table

Digestive tracts (N = 53)				
Prey type	Number of samples			Percentage of identifications relying on bones
	Otoliths	Bones	Either	
Clupeids	5	3	5	0
Flatfish	9	4	9	0
Gadids	27	15	27	0
Lumpsuckers	4	8	8	50
Salmonids	3	5	6	50
Sandeels	33	17	33	0

Common seal faeces (N = 503)				
Prey type	Number of samples			Percentage of identifications relying on bones
	Otoliths	Bones	Either	
Clupeids	98	134	155	37
Dragonets	2	4	6	67
Flatfish	37	30	48	23
Gadids	84	67	112	25
Sandeels	361	338	383	6

Grey seal faeces (N = 243)				
Prey type	Number of samples			Percentage of identifications relying on bones
	Otoliths	Bones	Either	
Catfish	1	11	11	91
Clupeids	3	6	8	63
Dragonets	6	8	10	40
Flatfish	28	14	37	24
Gadids	113	72	133	15
Lumpsuckers	1	9	9	89
Sandeels	97	79	110	12

All faeces (N = 823, groups occurring in < 10 samples excluded)				
Prey type	Number of samples			Percentage of identifications relying on bones
	Otoliths	Bones	Either	
Catfish	1	14	14	93
Clupeids	103	141	166	38
Dragonets	8	12	16	50
Flatfish	68	49	90	24
Gadids	210	147	262	20
Lumpsuckers	1	15	15	93
Sandeels	519	477	561	8

straightforward and misidentification at this level should be negligible. Identification to species obviously requires greater discrimination and affords greater opportunity for error.

Identification using skeletal remains other than otoliths

In digestive tracts, most otoliths and many bones were intact. Of the most commonly occurring prey groups, clupeids, flatfish (Heterosomata), gadids and sandeels (Ammodytidae) were all detected more frequently from otoliths than from bones, and the overall frequency of occurrence was not increased by identifying bones. However, salmonid and lumpsucker (*Cyclopterus lumpus*) bones were found more frequently than otoliths, and use of bones increased the frequency of detection (see Table VI).

In faeces (Table VI), otoliths sometimes showed obvious signs of erosion but most were identifiable. Bones were often fragmented, but many fragments were identifiable. The frequency of detection of all the common prey groups was increased by using bones in addition to otoliths, particularly for fish with small or fragile otoliths. Some fish were identified primarily from a single type of bone: catfish (*Anarhichas* sp.) from teeth; dragonets (*Callionymus lyra*) from preopercular spines; lumpsuckers from denticles. Rajids (which lack otoliths) were identified from denticles in four scats.

Discussion

Digestive tract samples were obtained incidentally, and we therefore have little information on the practical problems involved in collecting samples. Anecdotal information from local fishermen did, however, support the observation that seals often sink when shot (e.g. Fiscus & Baines, 1966); even when shot very close to land, carcasses were difficult to recover. Lavaging of stomach contents (e.g. Antonelis *et al.*, 1987), which offers a non-destructive alternative, has not been widely applied to seals.

Although sampling effort was not uniform across all seasons, it was apparent that faeces could not be obtained all year round at all sites. This reflects seasonal changes in the use of sites by seals. Thus, for example, most grey seal breeding sites in Orkney are deserted during the summer (McConnell, 1985), and common seals in Orkney use different haul-out sites at different times of year (Thompson, 1989). Clearly, it is important to have information on seasonal movements of seal populations in order to obtain year-round information on diet.

Common seal faeces were obtained in the Moray Firth throughout 1988 but, although more than two-thirds of sampling attempts yielded at least one sample, mean sample size was low. Samples were more readily obtained in the summer months. An obvious (but by no means certain) method of increasing sampling efficiency would be to sample only when large numbers of seals are present at a site.

Examination of digestive tract samples from both species indicated that, although many had empty stomachs, most contained recognizable food remains in some part of the tract. Therefore, to maximize sampling efficiency, it is suggested that the contents of the entire digestive tract should always be examined in dietary studies. If lavaging is to be attempted, useful additional information could be obtained by sampling faecal material from the colon.

In digestive tract samples, identification of bones resulted in no increase in frequency of detection for most prey groups, reflecting the good state of preservation of otoliths. The two groups (lumpsuckers, salmonids) for which this was not the case ironically both have relatively

friable skeletons. However, lumpsucker otoliths are very small and, at least in the case of seals feeding at salmon nets, salmon heads may not always be ingested (see, e.g. Rae & Shearer, 1965).

In faeces many bones were broken or fragmented, but use of bones significantly increased the rate of detection for most fish groups. We have not here taken identification to species level: small sample sizes would then make comparisons difficult in any case. The precision of identification of both otoliths and bones depends on familiarity with reference material and as such can be continually improved.

No adequate keys exist for skeletal elements other than otoliths in dietary remains of piscivores, although some authors have demonstrated the value of particular bones for identifying certain prey species for particular predators (Wise, 1980; Hansel *et al.*, 1988). Different fish are represented in seal scats by different bones, and it may therefore be inappropriate to concentrate entirely on a single type of bone (e.g. jaw or opercular bones) for all species. However, a given prey species may be reliably recognized from a single skeletal element.

The question arises as to whether use of bones in addition to otoliths would significantly alter estimates of diet composition in terms of fish biomass. Both bones and otoliths can be measured to estimate fish size (e.g. Casteel, 1976; Jobling & Breiby, 1986; Prime & Hammond, 1987; Hansel *et al.*, 1988). The extent to which otoliths of a given fish species are reduced in size is quantifiable using captive feeding experiments (e.g. Prime & Hammond, 1987; Harvey, 1989), although if the seals are inactive, passage rates may be reduced and digestion of otoliths increased (see Harvey, 1989). Although captive feeding experiments can be used to quantify the proportion of otoliths which are totally digested (e.g. da Silva & Neilson, 1985; Murie & Lavigne, 1986), obviously no meaningful correction factor can be applied when no otoliths are present in a field sample.

Some information is available on fish size-bone size relationships, e.g. in the archaeological literature (e.g. Casteel, 1976; Wheeler & Jones, 1976), but reductions in bone size and loss of bones during digestion have not been quantified.

Measurements on selected bones in seal faeces and digestive tracts are likely to be useful for improving estimates of diet composition, particularly for species known to be under-represented from otoliths. Further work is needed, however, to establish appropriate fish size-bone size measurement for the targeted prey species, and to quantify size reduction and loss of bones passing through seal digestive tracts.

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REFERENCES

- Antonelis, G. A., Lowry, M. S., Demaster, D. P. & Fiscus, C. H. (1987). Assessing northern elephant seal feeding habits by stomach lavage. *Mar. Mamm. Sci.* 3: 308–322.
- Bigg, M. A. & Fawcett, I. (1985). Two biases in diet determination of northern fur seals (*Callorhinus ursinus*). In *Marine mammals and fisheries*: 284–291. Beddington, J. R., Beverton, R. J. H. & Lavigne, D. M. (Eds). London: George Allen & Unwin.
- Bigg, M. A. & Perez, M. A. (1985). Modified volume: a frequency-volume method to assess marine mammal food habits. In *Marine mammals and fisheries*: 277–283. Beddington, J. R., Beverton, R. J. H. & Lavigne, D. M. (Eds). London: George Allen & Unwin.

- Breiby, A. (1985). *Otolitter fra saltvannfisker i nord-norge. Tromsø. Naturvitenskap nr. 45. Tromsø: Universitetet i Tromsø, Institutt for museumvirksomhet.*
- Brodeur, R. D. (1979). *Guide to otoliths of some Northwest Atlantic Fishes.* Massachusetts: National Marine Fisheries Service, Northeast Fisheries Center, Woods Hole Laboratory, Woods Hole.
- Casteel, R. W. (1976). *Fish remains in archaeology and paleo-environmental studies.* London & New York: Academic Press.
- Corbet, G. B. & Southern, H. N. (Eds) (1977). *The handbook of British mammals.* 2nd edn. Oxford: Blackwell.
- da Silva, J. & Neilson, J. D. (1985). Limitations of using otoliths recovered in scats to estimate prey consumption by seals. *Can. J. Fish. Aquat. Sci.* **42**: 1439–1442.
- Fiscus, C. H. & Baines, G. A. (1966). Food and feeding behavior of Steller and California sea-lions. *J. Mammal.* **47**: 195–200.
- Ford, E. (1937). Vertebral variation in teleost fish. *J. mar. biol. Ass. U.K.* **22**: 1–60.
- Hansel, H. C., Duke, S. D., Lofy, P. T. & Gray, G. A. (1988). Use of diagnostic bones to identify and estimate original lengths of ingested prey fishes. *Trans. Am. Fish. Soc.* **117**: 55–62.
- Härkönen, T. J. (1986). *Guide to the otoliths of the bony fishes of the northeast Atlantic.* Hellerup, Denmark: Danbiu.
- Härkönen, T. J. (1987). Seasonal and regional variations in the feeding habits of the harbour seal, *Phoca vitulina*, in the Skagerrak and the Kattegat. *J. Zool., Lond.* **213**: 535–543.
- Härkönen, T. J. (1988). Food-habitat relationship of harbour seals and black cormorants in Skagerrak and Kattegat. *J. Zool., Lond.* **214**: 673–681.
- Harvey, J. T. (1989). Assessment of errors associated with harbour seal (*Phoca vitulina*) faecal sampling. *J. Zool., Lond.* **219**: 101–111.
- Jobling, M. (1987). Marine mammal faecal samples as indicators of prey importance—a source of error in bioenergetics studies. *Sarsia* **72**: 255–260.
- Jobling, M. & Breiby, A. (1986). The use and abuse of fish otoliths in studies of feeding habits of marine piscivores. *Sarsia* **71**: 265–274.
- Kenyon, K. W. (1956). Food of fur seals taken on St Paul Island, Alaska, 1954. *J. Wildl. Mgmt* **20**: 214–215.
- McConnell, B. J. (1985). Seals in Orkney. *Proc. R. Soc. Edinb.* **87B**: 95–104.
- McConnell, B. J., Prime, J. H., Hiby, A. R. & Harwood, J. (1984). Grey seal diet. In *Interactions between grey seals and UK fisheries*: 148–183. S.M.R.U. Report on research conducted for the Department of Agriculture and Fisheries Scotland by the Natural Environment Research Council's Sea Mammal Research Unit. Cambridge: N.E.R.C.
- Mujib, K. A. (1967). The cranial osteology of the Gadidae. *J. Fish. Res. Bd Can.* **24**: 1315–1375.
- Murie, D. J. & Lavigne, D. M. (1986). Interpretation of otoliths in stomach content analyses of phocid seals: quantifying fish consumption. *Can. J. Zool.* **64**: 1152–1157.
- Norden, C. R. (1961). Comparative osteology of representative salmonid fishes, with particular reference to the grayling (*Thymallus arcticus*) and its phylogeny. *J. Fish. Res. Bd Can.* **18**: 679–791.
- Pikharev, G. A. (1946). The food of the seal *Phoca hispida*. *Fish. Res. Bd Can. (Transl. Ser.)* **150**: 1957.
- Prime, J. H. & Hammond, P. S. (1985). The diet of grey seals in the North Sea assessed from faecal analysis. In *The impact of grey seals on North Sea resources*: 84–99. Hammond, P. S. & Harwood, J. (Eds). Report to the Commission of the European Communities on Contract ENV 665 UK(H). Cambridge: N.E.R.C.
- Prime, J. H. & Hammond, P. S. (1987). Quantitative assessment of gray seal diet from fecal analysis. In *Approaches to marine mammal energetics*: 165–182. Huntley, A. C., Costa, D. P., Worthy, G. A. J. & Castellini, M. A. (Eds). Lawrence, Kansas: Society for Marine Mammalogy.
- Rae, B. B. (1960). Seals and Scottish fisheries. *Mar. Res.* **1960** (2): 1–39.
- Rae, B. B. (1968). The food of seals in Scottish waters. *Mar. Res.* **1968** (2): 1–23.
- Rae, B. B. (1973). Further observations on the food of seals. *J. Zool., Lond.* **169**: 287–297.
- Rae, B. B. & Shearer, W. M. (1965). Seal damage to salmon fisheries. *Mar. Res.* **1965** (2): 1–39.
- S.M.R.U. (1988). *Multispecies fishery assessment in the North Sea: estimation of mortality caused by marine mammals.* Final report on Contract with DGXIV-B-1 of the Commission of the European Communities. Cambridge: Sea Mammal Research Unit, Natural Environment Research Council.
- Thompson, P. M. (1989). Seasonal changes in the distribution and composition of common seal (*Phoca vitulina*) haul-out groups. *J. Zool., Lond.* **217**: 281–294.
- Wheeler, A. & Jones, A. (1976). Fish remains. In *Excavations on Fuller's Hill, Great Yarmouth*: 208–224. Rogerson, A. (Ed.). East Anglian Archaeology Report No. 2. Norfolk: Norfolk Archaeological Unit.
- Wise, M. H. (1980). The use of fish vertebrae in scats for estimating prey size of otters and mink. *J. Zool., Lond.* **192**: 25–31.

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Yellow-necked mice *Apodemus flavicollis* at Woodchester Park, 1968–1989

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The exceptional population of Yellow-necked mice *Apodemus flavicollis* occurring at Woodchester Park, Gloucestershire was first mentioned by Yalden (1971), who noted that on the grid studied there each June by students of Manchester University, *A. flavicollis* was twice as numerous as *A. sylvaticus*; generally, the wood mouse *Apodemus sylvaticus* is much more widespread and much more abundant in Britain than its congener. This abundance persisted to 1975 (Montgomery, 1976), and allowed the first substantial comparison of the ecology of the two *Apodemus* species in sympatry in Britain to be undertaken (Montgomery, 1977). In reviewing his own research and integrating it with the student studies, Montgomery (1985) pointed out that both the proportion and the absolute numbers of *A. flavicollis* on the main study grid declined after 1975. In this note, we bring the record for the main grid up to date, and also review the status of the species in the wider area of Woodchester Park.

Study area and methods

The main study grid, immediately below the field centre at Woodchester Park (Nat. Grid Ref. SO 811012), is a steeply sloping area of 0.77 ha (an effective trapping area, including boundary strips half a home range wide, of about 1.73 ha; Yalden, 1971). It is in mixed deciduous woodland, principally of *Fraxinus*, *Fagus* and *Taxus*, part of a continuous cover of woodland along the steep slopes of both sides of the valley. Much of the deciduous woodland has, however, been felled in the last 25 years, to be replaced by *Larix*, *Fagus*, *Thuja* and other commercial species. A fuller description of the valley is given by Askew & Yalden (1985).

In the early years, the size of the grid and the number of traps varied somewhat (Montgomery, 1976), but has been constant at 96 traps set at 10 m intervals (8 × 12 rows) since 1976. This is Grid M of Montgomery (1977, 1985). In June 1989, we laid 10 trap lines, each of 10 pairs of traps at 20 m intervals, in various sites around the valley; these sites were chosen, mostly, because they yielded *A. flavicollis* in earlier years. They were each set for four nights, as was the main grid. Individuals were marked by fur clipping; in the following analysis, the numbers of individuals handled at each site (rather than calculated populations) are used, for ease of comparison over the years. This procedure may underestimate the presence of bank voles *Clethrionomys glareolus*, which seem somewhat more 'trap-shy' than the *Apodemus* spp. (Montgomery, 1985).

Results

Main grid. In the years 1968–1975, *A. flavicollis* was generally twice as numerous as *A. sylvaticus*, and in six out of eight years was the most numerous woodland rodent (*C. glareolus* outnumbered it in 1968 and 1971). Numbers of *A. sylvaticus* and *A. flavicollis* were positively related, however